

Molecular docking and ADMET analysis of coenzyme Q10 as a potential therapeutic agent for Alzheimer's disease

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Abstract

Alzheimer's disease (AD) is a neurodegenerative disorder characterized by cognitive decline, synaptic dysfunction, and neuroinflammation, with oxidative stress playing a crucial role. Coenzyme Q10 (CoQ10), known for its antioxidant properties, has been proposed as a potential therapeutic agent due to its ability to mitigate oxidative damage and maintain mitochondrial integrity. This study investigates the molecular interactions of CoQ10 with key proteins involved in AD pathology—glycogen synthase kinase-3 β (GSK-3 β), protein kinase B (PKB), and phosphoinositide 3-kinase (PI3K)—through molecular docking methods. Results indicate that CoQ10 exhibits the strongest binding affinity with GSK-3 β , potentially reducing tau protein phosphorylation, a hallmark of AD. ADMET analysis further supports CoQ10's drug-like properties, with a favorable absorption and safety profile, although limited blood-brain barrier permeability poses a challenge. This *in silico* study highlights CoQ10's therapeutic potential in addressing neuroinflammation and oxidative stress in AD, warranting further *in vitro* and *in vivo* validation.

Keywords: Alzheimer's disease, coenzyme Q10, molecular docking, oxidative stress, neuroinflammation

Introduction

Alzheimer's disease (AD) is a chronic, progressive neurodegenerative condition predominately affecting the elderly, which causes substantial deficits in memory, cognitive deterioration, and synaptic connections difficulties [1]. It represents 60-80% of global dementia cases, and despite numerous research initiatives, no specific cure has been uncovered [2]. The condition known as Alzheimer's pathogenesis is characterized by two primary features: exogenous amyloid-beta (A β) plaques and intracellular neurofibrillary knots that consist of hyperphosphorylated tau proteins. For these characteristics, two essential patho-

genic mechanisms—neuroinflammation and oxidative stress—are important to the disease's development [3]. Neuroinflammation is the continuous activation of microglia and astrocytes, which produce pro-inflammatory cytokines that exacerbate damage to the neurons [4]. Oxidative stress is caused by an imbalance between the formation of reactive oxygen species (ROS) and the brain's defense against antioxidants, which causes damage to lipids, proteins, and genetic material [5].

Mitochondrial dysfunction has grown into an important contributor to oxidative stress in the development of AD [6]. Mitochondria, as the essential suppliers of cellular energy and ROS, contribute to increased damage from oxidation and neuroinflammation when malfunctioning. Coenzyme Q10 (CoQ10) is a possible chemical substance in the fight against oxidative stress, functioning as a naturally occurring antioxidant and a crucial component of the chain of electron transport in the mitochondria. CoQ10 works both in cellular production of energy and in neutralizing free radicals, therefore preserving neurons from damage caused by oxidation and protecting mitochondrial integrity. Additionally, CoQ10 has demonstrated neuroprotective properties in numerous neurodegenerative

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conditions, which include Parkinson's and Huntington's diseases, by lowering oxidative damage and inhibiting caspase pathways [7]. The neuroprotective impact of CoQ10 in the progression of Alzheimer's has been found to be connected to the ability to regulate important signaling pathways related to surviving cells and inflammation. The PI3K/Akt pathway plays an important role in improving neuronal life expectancy through the regulation of apoptosis, autophagy, respectively, and the inflammatory response. Three essential proteins in this pathway—glycogen synthase kinase-3 beta (the transcription factor GSK), protein kinase B, also known as PKB/Akt, and phosphoinositide 3-kinase (PI3K)—are particularly noteworthy in the study of Alzheimer's research. GSK-3 β contributes to the degradation of tau proteins, which produces neurofibrillary tangles, while PI3K and Akt perform protective roles by enhancing cell survival and preventing apoptosis. The process of deregulation of this pathway has been as-

sociated with the development of AD and increased susceptibility to damage from oxidation and inflammation [8]. Using a molecular docking method, this research is aimed at investigating the potential of CoQ10 to mitigate oxidative stress and neuroinflammation in AD, with particular emphasis on its connections with GSK-3 β , PKB/Akt, and PI3K. An effective way to investigate possible relationships between CoQ10 and target proteins, as well as potential regulation of the PI3K/Akt pathway, is by using the computational methodology of molecular docking. By analyzing CoQ10's impact on the proteins that signal associated with the major pathogenic mechanisms of AD, we expect to contribute molecular insights into the drug's potential. By comprehension of these molecular interactions, CoQ10 may become an alternative therapeutic intervention candidate in AD, providing a new approach to control neuroinflammation and oxidative stress. Beside that in the animal model, CoQ10 has been shown to have

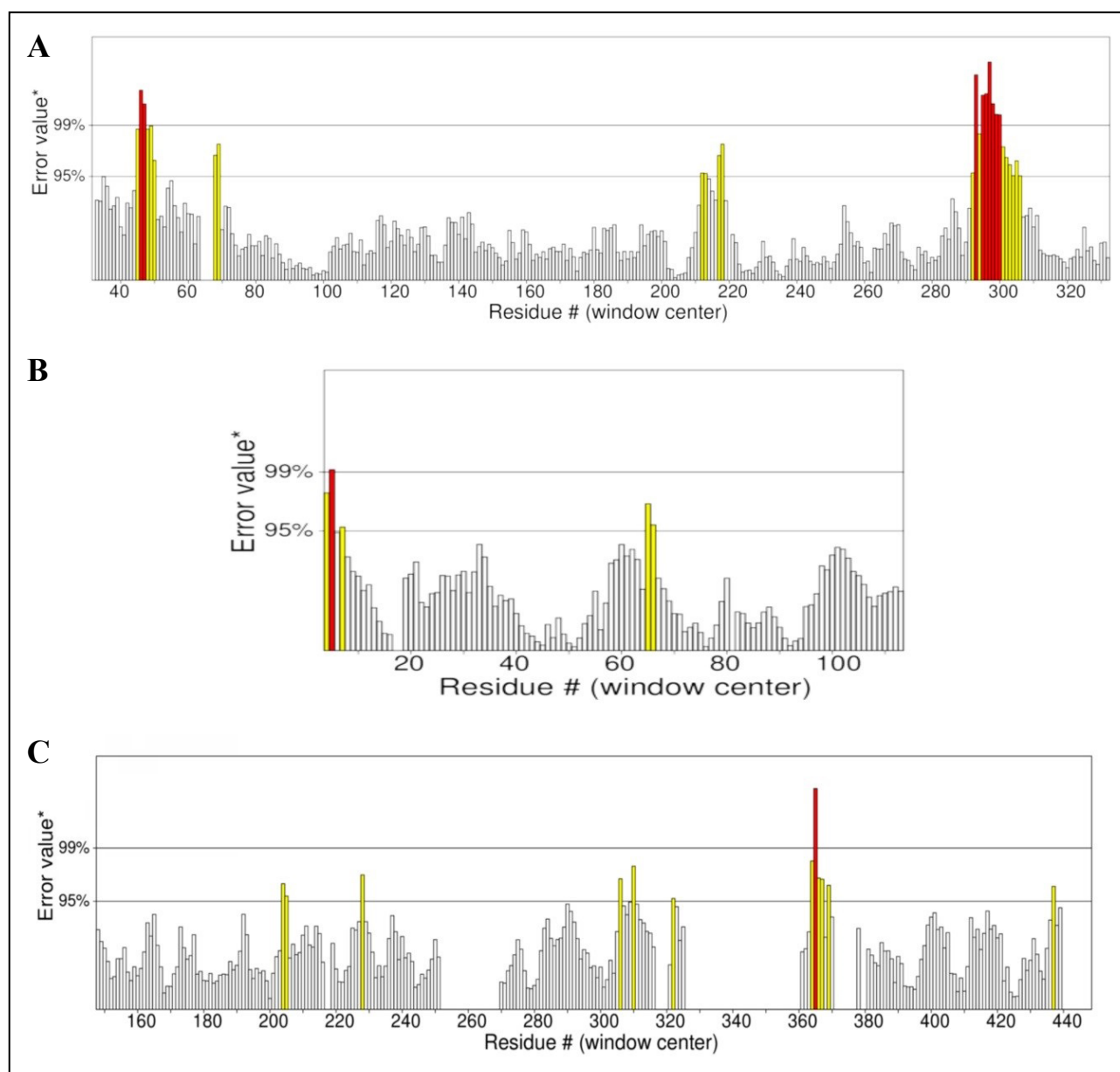


Figure 1. Error values distribution over all residues. The overall quality factor of GSK-3 β (A), PKB (B), and PI3K (C).

evidence of decreasing amyloid beta levels and formation of plaque. In animals, this explains why it may play a prominent role in mitigating AD pathology [9, 10].

Materials and methods

Molecular docking study

Computer-based tools were used to visualize the protein and ligand capabilities. These tools are very effective in molecular docking and save time [11]. Various online tools and software were used for this study, such as PyRx 0.8, ChimeraX 1.8, BIOVIA Discovery Studio 2024 Client, PkCSM, Errat, and Pro-SA Web.

Ligand selection

For this study, CoQ10 was selected. CoQ10 is an influential antioxidant that is produced by our body inherently [12]. This coenzyme has significant antioxidant properties. This coenzyme can be used as an important bioactive compound to relieve the oxidation [13]. The antioxidant character of CoQ10 helps to remove oxidative stress from cells. Oxidative stress is mainly connected to AD [14, 15]. Moreover, some research has found that CoQ10 plays a significant role in decreasing amyloid- β plaques and tau protein tangles; these factors are connected to AD [15, 16]. Overall, this compound was thought to be perfect for the study.

Protein selection

CoQ10 interacts with a number of important proteins involved in IL-17-mediated neurological inflammation and oxidative stress in AD [17, 18]. The selected proteins are 1. phosphoinositide 3-kinase (PI3K) (PDB ID: 3APD), 2. protein kinase B (PKB) (PDB ID: 1UNQ), and 3. glycogen synthase kinase-3 β (GSK-3 β) (PDB ID: 1GNG). PI3K and PKB are parts of the PI3K and Akt signaling pathways. These proteins have a significant impact on neuroprotection and survival of cells [19]. Further, GSK-3 β is involved in tau protein phosphorylation [19].

Ligand preparation

From previous findings, CoQ10 (PDB ID: 521915) was

selected [20]. The structure of this compound was collected from Pubchem (<https://pubchem.ncbi.nlm.nih.gov/>, accessed on October 4, 2024) in SDF format for binding to selected proteins. To start with, the coenzyme was directly inserted in the PyRx software. Other software recommends the pdbqt format, but in PyRx, there is no need for the pdbqt file, because PyRx directly supports sdf files [21]. After that, energy was reduced of that compound for a perfect result. For docking, the compound was then converted into pdbqt format. Science PyRx has access to openable [22]. After performing docking, the specific conformation of this compound is selected and saved as a PDB file for further analysis.

Protein check

Errat

The accuracy and reliability of the proteins were detected by ERRAT (<https://saves.mbi.ucla.edu/>, accessed on October 4, 2024), an online program that analyzes the quality of a protein structure by examining non-bonded atom-atom interactions [23]. An overall quality factor greater than 90% shows a high quality model [24]. According to Errat, the overall quality factor of PI3K (PDB ID: 3APD) was 95.641% (Figure 1C), PKB (PDB ID: 1UNQ) was 95.327% (Figure 1B), and GSK-3 β (PDB ID: 1GNG) was 92.222% (Figure 1A). The overall quality factors of all proteins are above 90%, which means high-quality model. Based on Figure 1, the x-axis represents all residues and the y-axis represents the errors. The red lines show the higher error value, which is above 99% or only at 99%, and the yellow vertical lines also indicate the error value, but at a low level. Moreover, the two horizontal lines at 95% and 99% are the threshold lines.

ProSA-Web

Moreover, the ProSa-Web tool (<https://prosa.services.came.sbg.ac.at/prosa.php>, accessed on October 4, 2024) was used to check the proteins. It is a useful online tool that finds problems in three-dimensional protein structures. It also gives score and energy graphs to highlight faults in protein models and the results are shown in graphs (Figure 2 and Figure 3) [25]. The more negative value of the z-score represents a better protein model [26,

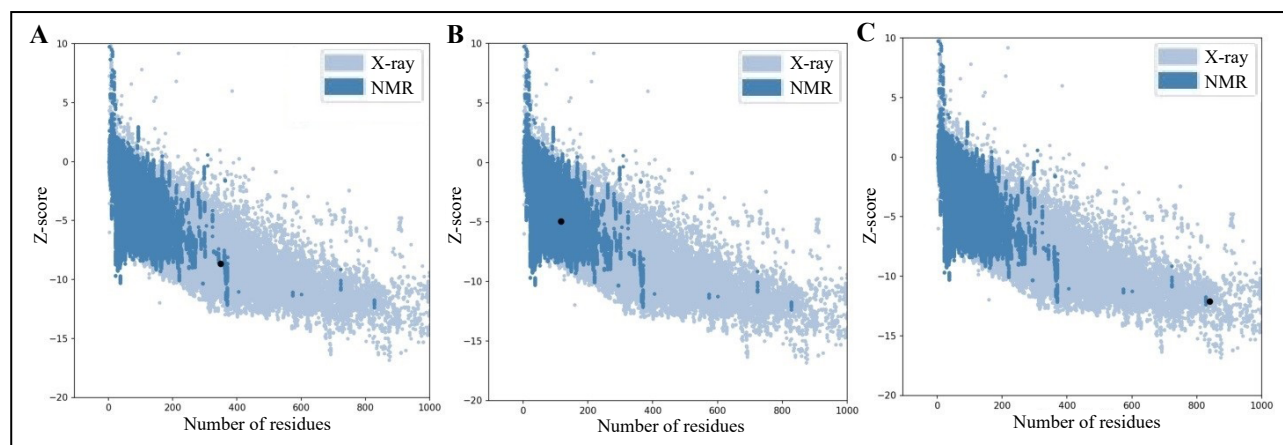


Figure 2. Z-score graphs for identified proteins, including GSK-3 β (A), PKB (B), and PI3K (C).

27].

According to Figure 2, the z-score for GSK-3 β is -8.68, which indicated that the protein is located in the NMR solved protein structure (Figure 2A); PKB is -4.97, which represented that the protein is present in the NMR solved protein structure (Figure 2B); and PI3K is -12.1, which showed that the protein structure is situating in X-ray solved protein structure (Figure 2C). The z-scores are represented as black dots in the graphs. Besides, the x-axis and y-axis of the energy plots (Figure 3) show the position of the amino acid sequences and the energy values. On the y-axis, positive values show the problematic regions and negative values represent the correct regions of a protein. In Figure 3, it is noticeable that GSK-3 β (Figure 3A) and PKB (Figure 3B) have more negative energy levels than PI3K (Figure 3C).

Hydrophobicity

The hydrophobicity graphs in Figure 4 show the hydrophobicity properties of amino acid residues in a protein sequence. The x-axis indicates the position of the amino acid residues and the y-axis represents the average hydrophobicity of each amino acid. Again, the yellow circles exhibit the hydrophobicity of each residue and the blue lines connect the yellow circles to help visualize the data point's position. The positive values above 0 display the hydrophobic areas and are found in the interior of the protein as well as in the transmembrane region. On the other hand, the negative values below 0 show the hydrophilic areas that are present in the interaction with other molecules.

Protein preparation

The identified proteins are prepared one by one using the UCSF ChimeraX software. It is a reliable software for preparing a protein for molecular docking studies [28]. First, a protein's pdb file is inserted in the software. Next, the heteroatoms are deleted, and one chain is selected for the protein, which has two or more protein chains. After that, the Dock Prep option is selected for full preparation. In this section, H-bonds, charges and missing residues are added to the protein [29]. After completing these steps,

the newly prepared protein is saved as a pdb file. Furthermore, the active sites of the proteins are identified using Discovery Studio [30].

Protein-ligand binding

Before binding proteins to the specific compounds, their active sites are determined using Discovery Studio. This software is very useful to quickly identify the active sites [31]. For GSK-3 β (PDB ID: 1GNG), ARG96, ARG180, LYS205, ARG209, ASN213, VAL214, and VAL208 are the amino acids as active sites. Again, amino acids including SER0, MET1, SER2, HIS13, LYS14, GLY16, GLU17, TYR18, ILE19, ARG23, ARG25, LEU52, ASN53, PHE55, and ARG86 are selected as active binding sites for PKB (PDB ID: 1UNQ). TRP212, LYS288, ILE831, LYS833, ASP836, ILE879, GLU880, ILE881, VAL882, THR887, ASP946, ARG947, HIS948, ASN951, MET953, ILE963, ASP964, and ILE968 are identified for PI3K (PDB ID: 3APD). During molecular docking, ligands do not always bind to the identified active sites of a protein. This situation is known allosteric regulation [32]. For perfect binding, the protein grid box was maximized. As a result, whole proteins become selected and ligands can bind to the perfect site. First, a specific protein and ligand were inserted into PyRx for molecular docking. Next, different positions and dimensions were set for the grid box for maximization due to each protein. The grid box was fixed into center_x = 31.1276, center_y = -0.0207, center_z = 26.3320 and dimension_x = 81.9806, dimension_y = -63.7440, dimension_z = 59.5978 for 3APD. For 1GNG, the grid box was set into center_x = 108.3264, center_y = 55.8664, center_z = 35.0965 and dimension_x = 36.7377, dimension_y = 30.3748, dimension_z = 34.9006. And the box was selected into center_x = 21.7185, center_y = 14.4762, center_z = 9.7653 and dimension_x = 40.8121, dimension_y = 38.0033, dimension_z = 49.1876 for 1UNQ. PyRx has accessibility to Auto-dock Vina to perform docking [33]. After the docking is finished, the significant conformation for the selected compound was saved as a pdb file for each protein-ligand binding. During selection, the binding energy for each conformation was determined, and the conformation

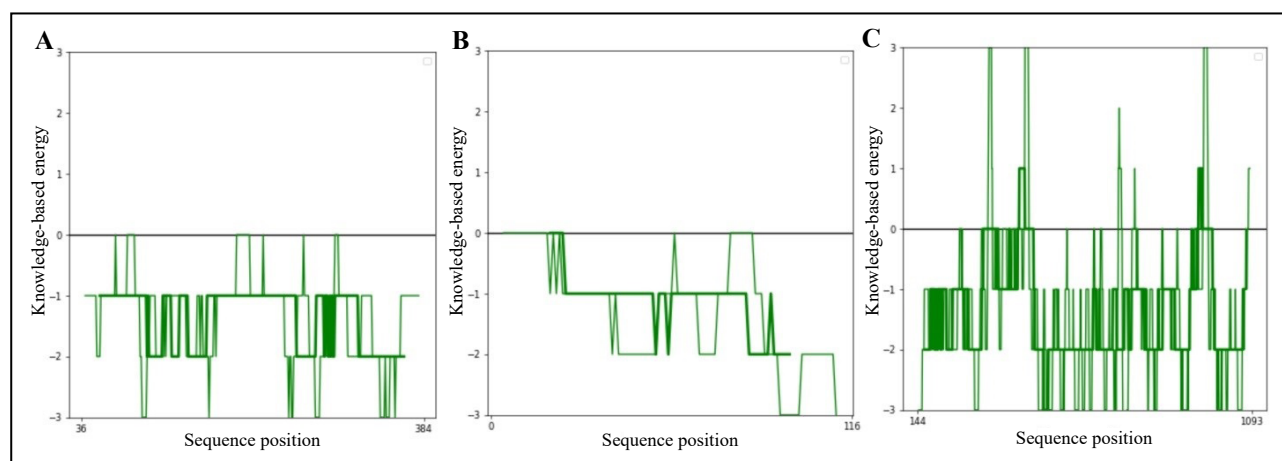


Figure 3. Z-score graphs for identified proteins, including GSK-3 β (A), PKB (B), and PI3K (C).

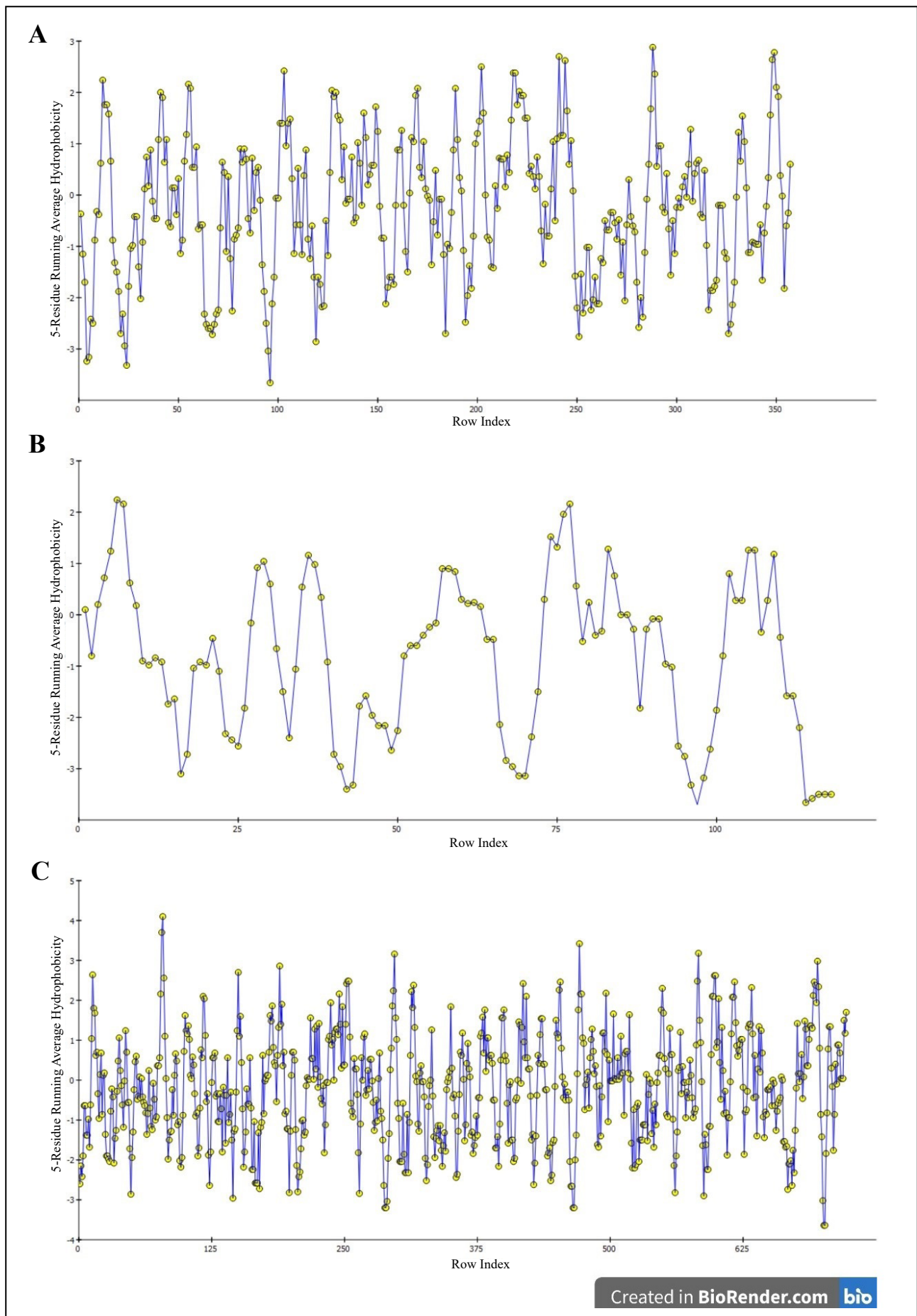


Figure 4. Hydrophobicity graphs for GSK-3 β (A), PKB (B), and PI3K (C).

Table 1. Detailed ADMET screening for docking compound.

Property	Model name	Predicted value	Unit
Absorption	Water solubility	-3.255	Numeric (log mol/L)
Absorption	Caco-2 permeability	1.296	Numeric (log Papp in 10 ⁻⁶ cm/s)
Absorption	Intestinal absorption (Human)	91.871	Numeric (% Absorbed)
Absorption	Skin permeability	-2.735	Numeric (log Kp)
Absorption	P-glycoprotein substrate	No	Categorical (Yes/No)
Absorption	P-glycoprotein I inhibitor	No	Categorical (Yes/No)
Absorption	P-glycoprotein II inhibitor	Yes	Categorical (Yes/No)
Distribution	VDss (Human)	-0.974	Numeric (log L/kg)
Distribution	Fraction unbound (Human)	0.146	Numeric (Fu)
Distribution	BBB permeability	-0.961	Numeric (log BB)
Distribution	CNS permeability	-1.176	Numeric (log PS)
Metabolism	CYP2D6 substrate	No	Categorical (Yes/No)
Metabolism	CYP3A4 substrate	Yes	Categorical (Yes/No)
Metabolism	CYP1A2 inhibitor	No	Categorical (Yes/No)
Metabolism	CYP2C19 inhibitor	No	Categorical (Yes/No)
Metabolism	CYP2C9 inhibitor	No	Categorical (Yes/No)
Metabolism	CYP2D6 inhibitor	No	Categorical (Yes/No)
Metabolism	CYP3A4 inhibitor	No	Categorical (Yes/No)
Excretion	Total clearance	1.345	Numeric (log mL/min/kg)
Excretion	Renal OCT2 substrate	No	Categorical (Yes/No)
Toxicity	AMES toxicity	No	Categorical (Yes/No)
Toxicity	Max. tolerated dose (Human)	0.225	Numeric (log mg/kg/day)
Toxicity	hERG I inhibitor	No	Categorical (Yes/No)
Toxicity	hERG II inhibitor	No	Categorical (Yes/No)
Toxicity	Oral rat acute toxicity (LD50)	2.344	Numeric (mol/kg)
Toxicity	Oral rat chronic toxicity (LOAEL)	3.667	Numeric (log mg/kg_bw/day)
Toxicity	Hepatotoxicity	No	Categorical (Yes/No)
Toxicity	Skin sensitization	No	Categorical (Yes/No)
Toxicity	<i>T. Pyriformis</i> toxicity	0.285	Numeric (log µg/L)
Toxicity	Minnow toxicity	-9.793	Numeric (log mM)

of a ligand with the lowest binding energy was selected [34]. Finally, the proteins and ligands were entered into the Discovery Studio tool for further analysis [35].

ADMET analysis

To understand the pharmacokinetics, such as absorption, distribution, metabolism, excretion, and toxicity of a drug, computer-based tools are used worldwide. The ADMET properties are strictly followed to verify the drug likeness properties for finding new drugs [36]. Compounds need to follow Lipinski's rules to be a drug. However, it is not necessary to follow Lipinski's rules. Some compounds

may not follow these rules and still be effective drugs, such as natural products [37]. The tool named pkCSM (<https://biosig.lab.uq.edu.au/pkcsm/prediction>, accessed on October 4, 2024) was used to analyze the ADMET properties. Table 1 shows the ADMET analyses and drug-like prediction of property for this significant chemical.

Results

Protein-to-ligand binding

First, Table 2 shows the binding energy of CoQ10 with

Table 2. Baseline characteristics of the data.

SN	Protein name	PDB ID	Chain	Mutation	Structure weight	Binding energy Coenzyme Q10
1	GSK-3β	1GNG	B	No	95.85 k DA	-7.2 kcal/mol
2	PKB	1UNQ	A	No	15.3 k DA	-4.6 kcal/mol
3	PI3K	3APD	A	No	111.02 k DA	-6.4 kcal/mol

GSK-3 β , which is -7.2 kcal/mol. The binding structure is shown more clearly in Figure 5. Figure 5A shows the 3D binding structure and Figure 5B presents the 2D diagram. In Figure 5B, we can observe that CoQ10 developed connections with seven amino acids of 1GNG, including ARG220, TYR221, LYS85, GLN185, LEU132, VAL70, and ALA83. There are four pi-alkyl bonds, two alkyl bonds, and one carbon-hydrogen bond present between the interactions of CoQ10 with GSK-3 β (Figure 5B).

Furthermore, Table 2 represents the binding affinity of CoQ10 with PKB, which is -4.6 kcal/mol. We can visualize their binding site in Figure 6. Figure 6A reveals the 3D binding structure between PKB and CoQ10. Figure 6B indicates the 2D structure. GLN79, TRP80, ASN54, LEU78, THR34, SER56, ASP32, GLY33, LEU110, ALA58, GLU114, and GLN113 are the amino acids with which the compound in Figure 6B interacts. Van der waals bonds, carbon-hydrogen bonds, alkyl, and pi-alkyl bonds are also visible in this figure.

Moreover, Table 2 exhibits the binding validity of CoQ10 with PI3K, which is -6.4 kcal/mol. Figure 7 provides the 3D configuration (Figure 7A) and 2D arrangement (Figure 7B) of PI3K with the chosen compound. In Figure 7B, it is noticeable that CoQ10 (compound) has developed the connections between the amino acids including, LEU823, TRP292, ARG277, HIS 295, ASP788, PRO866, and LEU865 of 3APD (protein). One conventional hydrogen bond, carbon-hydrogen bonds, alkyl, and pi-alkyl bonds are also observable in this figure.

Ramachandran plot analysis

The Ramachandran plot is the graphical representation that indicates the phi (ϕ) and psi (ψ) angles of the amino acids in a protein [38]. The x-axis exhibits the phi angle and the y-axis shows the psi angle [39]. The graph indicates distinctive regions where various phi and psi combi-

nations are available. The important regions are the left-handed alpha helix, the right-handed alpha helix, and the beta sheet.

In Figure 8, it is noticeable that amino acid residues of all proteins are heavily represented in the favorable regions. Besides, some residues exist outside of this region. According to Figure 8B, amino acids (green colored) are clustered in the favorable regions (red colored), which means that most of the residues are in stable conformations, and glycine residues (purple colored) show more flexibility. Again, Figure 8A and 8C exhibit that most of the residues are energetically present in the favorable regions, which indicate the stable conformation of these proteins, and glycine residues represent flexibility.

ADMET analysis for CoQ10

Absorption

According to Table 1:

1. Water solubility: This indicates the compound's solubility in water. The negative number indicates low solubility, which may affect its bioavailability when taken orally [40].
2. Caco-2 permeability: Finds out the rate at which a substance can enter the Caco-2 cell monolayer. The value greater than 1.0 is considered to have good permeability, which means the substance can be properly absorbed through the gut [41].
3. Intestinal absorption (human): This value indicates the percentage of a compound that is absorbed by the intestine. The high value means excellent absorption and is perfect for oral drugs [42].
4. Skin permeability: Displays the ability of that compound to enter into skin. More negative value means low skin permeability, but might be ideal for other routes [43].
5. P-glycoprotein substrate: indicates that the compound is not an activator for P-glycoprotein. As a result, this pro-

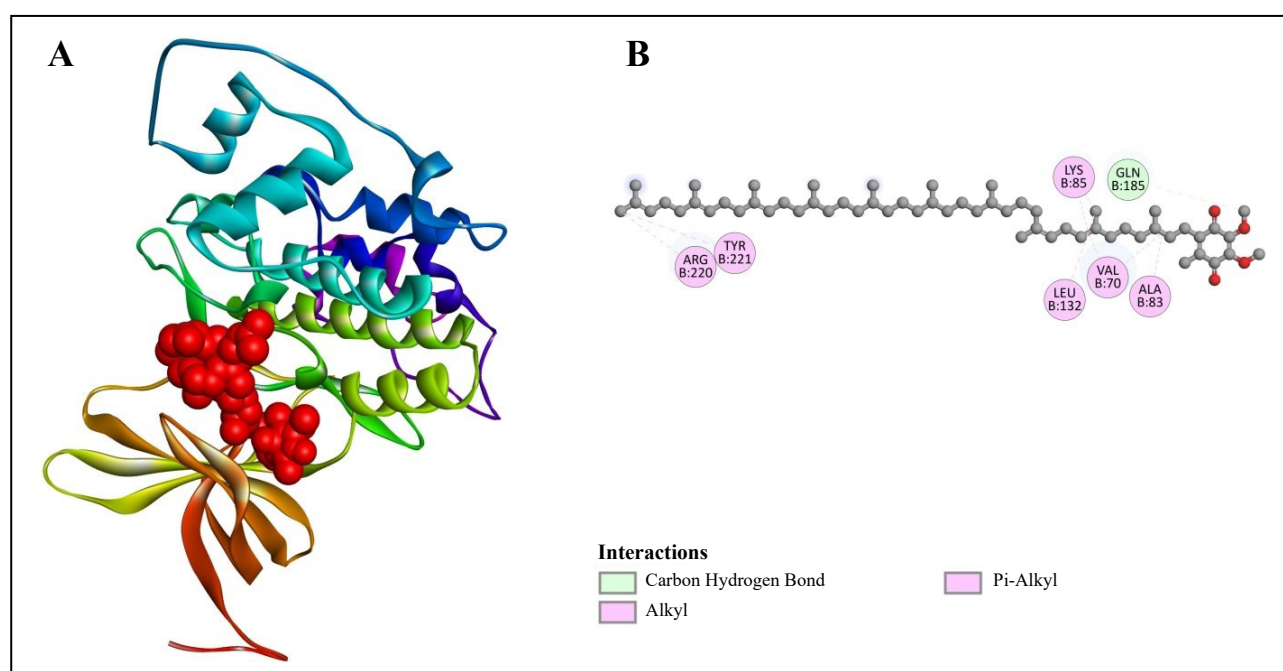


Figure 5. 3D (A) and 2D (B) structures of coenzyme Q10 binding with GSK-3 β .

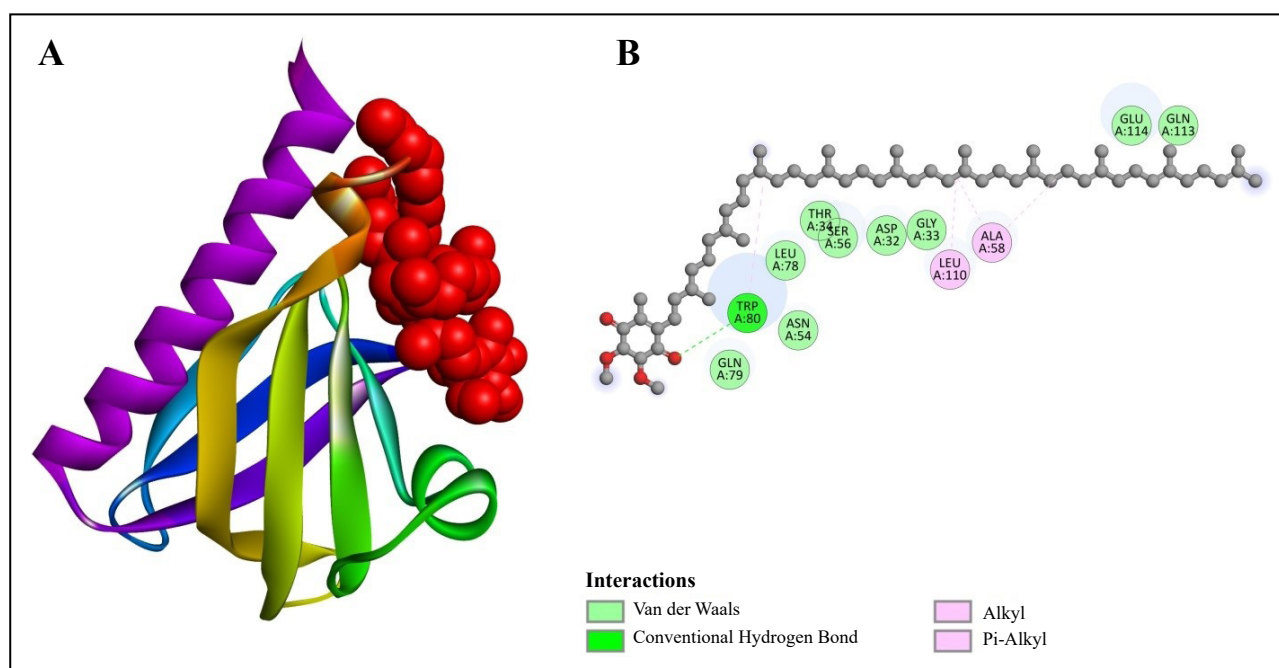


Figure 6. 3D (A) and 2D (B) structures of coenzyme Q10 binding with PKB.

tein cannot pump drugs to cells [44].

6. P-glycoprotein I inhibitor: The compound does not inhibit P-glycoprotein I, which shows that it will reduce drug-drug interaction [45].

7. P-glycoprotein II inhibitor: But inhibits P-glycoprotein II.

Distribution

Based on Table 1:

1. VDss (human): The Volume of distribution at steady state (VDss) exhibits how quickly a compound distributes throughout the body. The negative result states low distribution, suggesting that the drug stays in the plasma of blood [46].

2. Fraction unbound (human): Displays the fraction of the drug that is unbound and free for interaction with the target site. The lower value represents the higher binding with plasma protein [47].

3. BBB permeability: Shows the ability of a compound to cross the Blood-Brain-Barrier. The negative value represents low BBB permeability [48].

4. CNS permeability: This value helps to identify a compound's capability to enter into CNS tissues. Negative result is showing poor CNS activation [49].

Metabolism

As mentioned by Table 1:

1. CYP2D6 substrate: This result indicates that the compound cannot be broken down by the cytochrome P450 2D6 enzyme, which limits the possibility of certain metabolic interactions [50].

2. CYP3A4 substrate: The compound presents that it can be metabolized by CYP3A4 enzyme. It is a very important enzyme for drug metabolism [51].

3. CYP1A2 inhibitor: The compound shows that it does not create disturbance with the CYP1A2 enzyme.

4. CYP2C19 inhibitor: The identified substance is not an inhibitor for CYP2C19, which means it minimizes the drug-drug interactions [52].

5. CYP2C9 inhibitor: The compound indicates that it does not interfere in the metabolism of drugs that are processed by the CYP2C9 enzyme.

6. CYP2D6 inhibitor: Shows no inhabitation, which means decreases the risk of drug-drug connections.

7. CYP3A4 inhibitor: The compound does not inhibit CYP3A4.

Excretion

From the point of view of Table 1:

1. Total clearance: The total clearance value exhibits that the compound has a moderate clearance rate.

2. Renal OCT2 substrate: The result represents that the compound is not a substrate for renal organic cation transporter 2. As a result, the compound won't be affected while transporting through that pathway [53].

Toxicity

According to Table 1:

1. AMES toxicity: The compound has no AMES toxicity, which means it is not mutagenic and will not cause any genetic mutations [54].

2. Max. tolerated dose (human): 0.225 log mg/kg/day is the max dose of that substance for humans. This is the safe and helpful dose level according to the substance [55].

3. hERG I and II inhibitor: It does not inhibit hERG channels and decrease cardiac toxicity risk [56].

4. Oral rat acute toxicity (LD50): 2.344 mol/kg is the compound's acute toxicity level [57].

5. Oral rat chronic toxicity (LOAEL): 3.667 log mg/kg_{bw}/day is the level when the compound can create chronic toxicity [58].

6. Hepatotoxicity: It has no hepatotoxicity which results

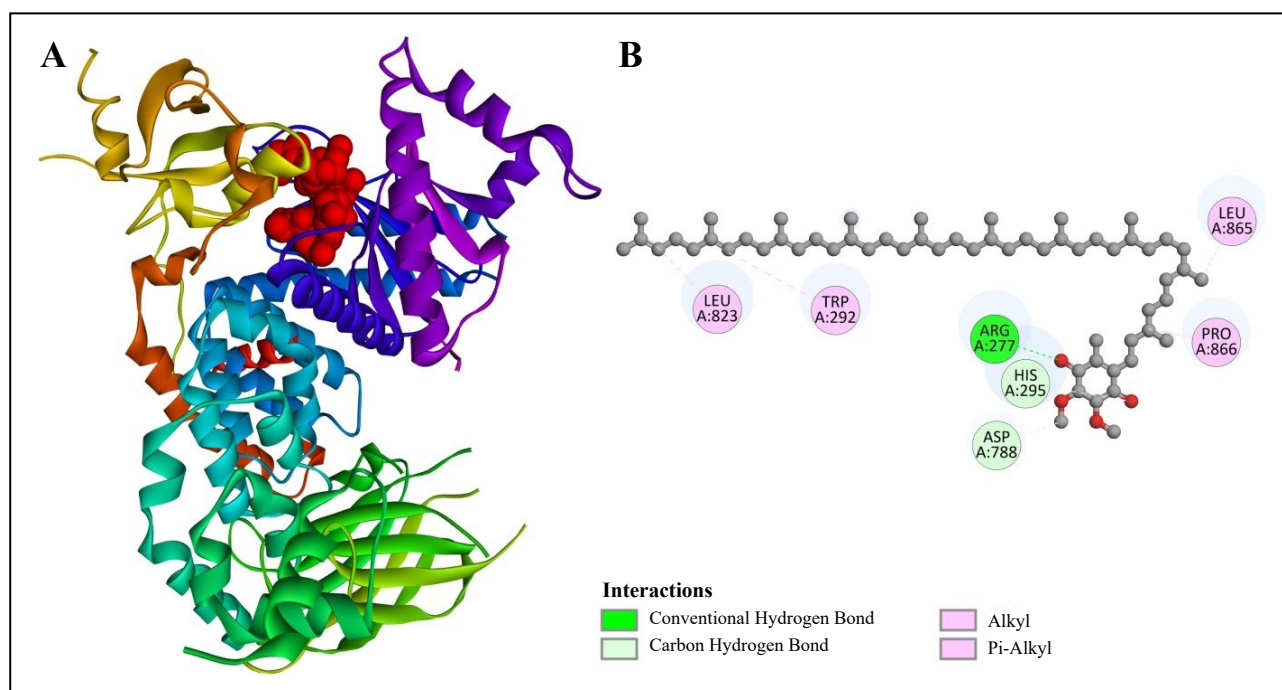


Figure 7. 3D (A) and 2D (B) structure of coenzyme Q10 binding with PI3K.

in no toxicity in the liver and ensures the overall safety [59].

7. Skin sensitization: The compound has no skin sensitization, which indicates it will not create an allergic reaction in the skin [60].

8. *T. pyriformis* toxicity: 0.285 log $\mu\text{g/L}$ shows that the compound can cause a low toxicity to *T. pyriformis* [61].

9. Minnow toxicity: The value is negative, which means low toxicity to fish [62].

Discussion

The present investigation focuses on the therapeutic properties of CoQ10 to find the possibilities to eradicate AD, based on molecular docking and ADMET analysis. The results provide significant information about the connection between CoQ10 and the proteins that are involved in oxidative stress and brain inflammasome [63]. These are the pathways that enhance AD. The molecular docking results exposed links between CoQ10 and the target proteins: GSK-3 β , PKB, and PI3K.

CoQ10 shows the highest binding affinity with GSK-3 β , which is -7.2 kcal/mol, and has developed connections with seven amino acids, including ARG220, TYR221, and LYS85. This strong interaction presents that CoQ10 can affect GSK-3 β activity, which is crucial for tau protein phosphorylation. Furthermore, the binding affinity of CoQ10 with PI3K is -6.4 kcal/mol with the interactions between the compound and amino acids such as LEU823, TRP292, and ARG277. These interactions with PI3K are involved in the PI3K/Akt signaling pathway, which plays a significant role in neuroprotection and cell survival. Additionally, CoQ10 exhibits the lower binding affinity with PKB, which is -4.6 kcal/mol, and the compound forms

connections with various amino acids, including GLN79, TRP80, and ASN54. These interactions may contribute to the PI3K/Akt pathway, potentially increasing the neuroprotective effects. Based on previous studies, these molecular interactions represent the mechanistic basis for the CoQ10 effects on antioxidant and neuroprotection [13-15]. The capability of CoQ10 to interact with these proteins highlights the compound's possibility in reducing oxidative stress and brain inflammation [64].

The ADMET analysis of CoQ10 revealed several favorable pharmacokinetic and toxicological properties [65]:

1. Absorption: CoQ10 expresses high intestinal absorption (91.871%) and good Caco-2 permeability (1.296), indicating excellent oral bioavailability. But it has low water solubility (-3.255 log mol/L), which may create challenges for formulation and delivery [40-42].

2. Distribution: The negative BBB permeability (-0.961) and CNS permeability (1.176) results present limited entrance into the central nervous system. This is a limitation for treating neurological disorders like AD [46-48].

3. Metabolism: CoQ10 is a substrate for CYP3A4 but not for CYP2D6. Further, it does not inhibit major CYP enzymes. These properties show the lower risk of drug-drug interactions. It is valuable for a patient who takes multiple drugs at a time [66].

4. Excretion: The compound showed a medium clearance rate, which is 1.345 log mL/min/kg, which indicates a balanced elimination profile.

5. Toxicity: CoQ10 published a favorable toxicity profile with no AMES toxicity, hepatotoxicity, or skin sensitization. It also showed no inhibition of hERG channels, which reduces the risk of cardiotoxicity. The maximum tolerated dose and chronic toxicity levels mean that it is safe for human use [54-56].

These ADMET properties establish the safety profile for

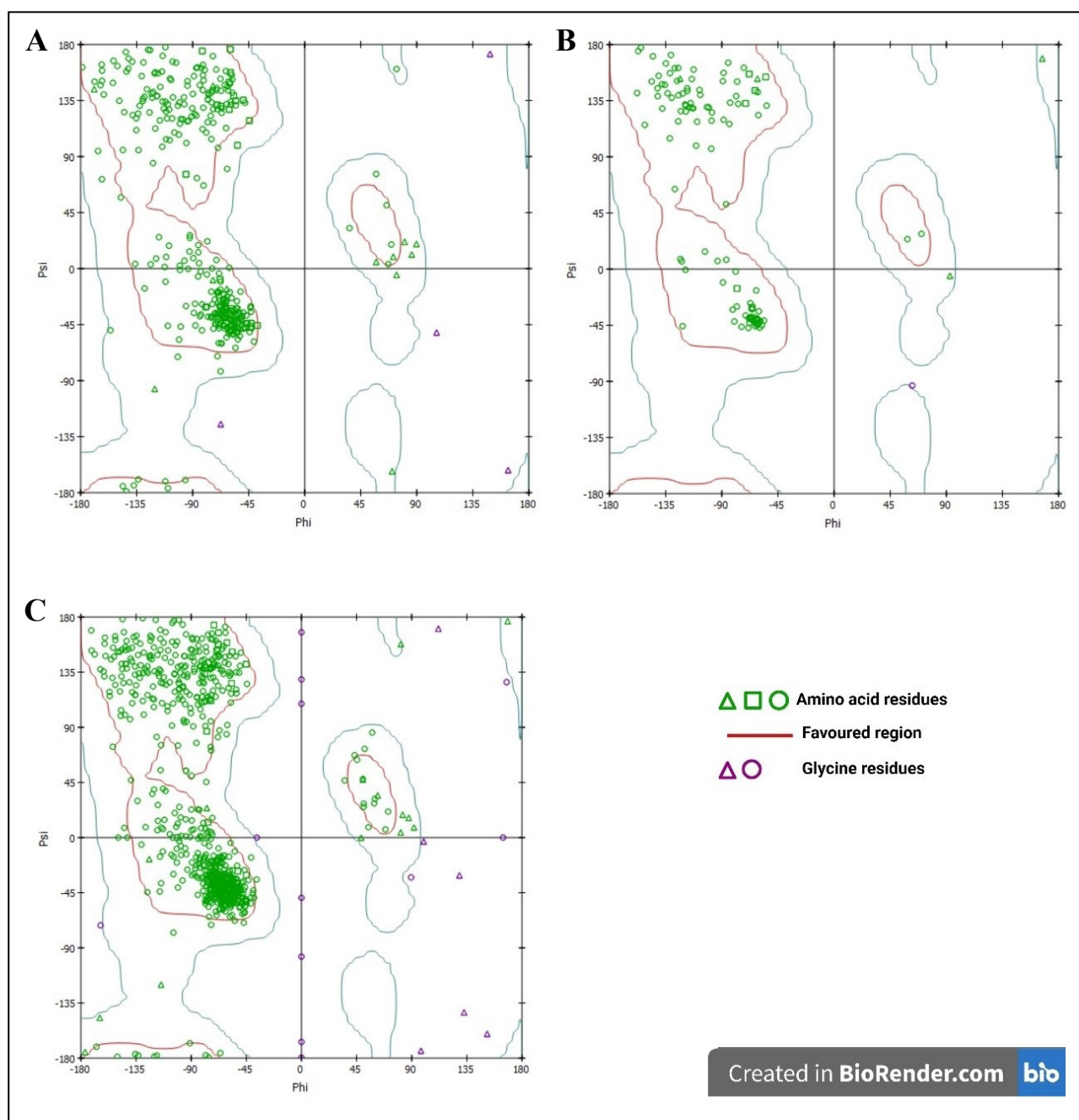


Figure 8. 3D (A) and 2D (B) structure of coenzyme Q10 binding with PI3K.

CoQ10 in clinical use and support its potential for long-term use in chronic conditions like AD.

The proper binding affinities between CoQ10 and the key proteins involved in AD pathology and its positive AD-MET result encourage further investigation on CoQ10 for its potential therapeutic properties for treating AD. The interactions with GSK-3 β show a better result that indicates CoQ10 may help in reducing tau protein phosphorylation, a hallmark of AD [19].

However, the poor BBB permeability of CoQ10 presents a significant challenge that needs to be focused on. Future research should place importance on developing the delivery systems and chemical modifications to enhance the CNS penetration of CoQ10 to increase its beneficial properties.

Additionally, this *in silico* study provides valuable data [67]. This information needs to be identified through *in vitro* and *in vivo* experiments. More studies are needed to find out the impacts of CoQ10 on tau phosphorylation, amyloid- β plaque formation, and neuroinflammation.

Conclusions

This study demonstrates the therapeutic potential of CoQ10 in mitigating key pathological mechanisms associated with AD, such as oxidative stress and neuroinflammation. Molecular docking results revealed strong interactions between CoQ10 and GSK-3 β , PKB, and PI3K, suggesting that CoQ10 may reduce tau phosphorylation

and enhance neuroprotection via the PI3K/Akt signaling pathway. ADMET analysis confirmed CoQ10's favorable pharmacokinetic profile, including high intestinal absorption and low toxicity, though its limited blood-brain barrier permeability remains a challenge for treating central nervous system disorders like AD.

Overall, this *in silico* investigation supports CoQ10 as a promising candidate for therapeutic development in AD. However, further experimental studies, including *in vitro* and *in vivo* models, are required to validate its efficacy in reducing amyloid- β plaques, tau tangles, and overall disease progression. Future research should also explore strategies to enhance CoQ10 delivery to the brain for more effective treatment outcomes.

Declarations

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References

- Kumar A, Sidhu J, Lui F, & Tsao JW. Alzheimer disease. *In: StatPearls*. 2024, edn.
- Holtzman DM, Morris JC, & Goate AM. Alzheimer's disease: the challenge of the second century. *Sci Transl Med*, 2011, 3(77): 77sr71. [[Crossref](#)]
- Vaiserman A, Koliada A, & Lushchak O. Neuroinflammation in pathogenesis of Alzheimer's disease: phytochemicals as potential therapeutics. *Mech Ageing Dev*, 2020, 189: 111259. [[Crossref](#)]
- Adamu A, Li S, Gao F, & Xue G. The role of neuroinflammation in neurodegenerative diseases: current understanding and future therapeutic targets. *Front Aging Neurosci*, 2024, 16: 1347987. [[Crossref](#)]
- Selkoe DJ, & Hardy J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol Med*, 2016, 8(6): 595-608. [[Crossref](#)]
- Wang X, Wang W, Li L, Perry G, Lee HG, & Zhu X. Oxidative stress and mitochondrial dysfunction in Alzheimer's disease. *Biochim Biophys Acta*, 2014, 1842(8): 1240-1247. [[Crossref](#)]
- Wyss-Coray T, & Rogers J. Inflammation in Alzheimer disease—a brief review of the basic science and clinical literature. *Cold Spring Harb Perspect Med*, 2012, 2(1): a006346. [[Crossref](#)]
- Beurel E, Grieco SF, & Jope RS. Glycogen synthase kinase-3 (GSK3): regulation, actions, and diseases. *Pharmacol Ther*, 2015, 148: 114-131. [[Crossref](#)]
- Beal MF. Coenzyme Q10 administration and its potential for treatment of neurodegenerative diseases. *Biofactors*, 1999, 9(2-4): 261-266. [[Crossref](#)]
- Butterfield DA, & Halliwell B. Oxidative stress, dysfunctional glucose metabolism and Alzheimer disease. *Nat Rev Neurosci*, 2019, 20(3): 148-160. [[Crossref](#)]
- Zhang B, Li H, Yu K, & Jin Z. Molecular docking-based computational platform for high-throughput virtual screening. *CCF Trans High Perform Comput*, 2022, 4(1): 63-74. [[Crossref](#)]
- Scarpelli M, Todeschini A, Rinaldi F, Rota S, Padovani A, & Filosto M. Strategies for treating mitochondrial disorders: an update. *Mol Genet Metab*, 2014, 113(4): 253-260. [[Crossref](#)]
- Mantle D, & Dybring A. Bioavailability of coenzyme Q(10): an overview of the absorption process and subsequent metabolism. *Antioxidants (Basel)*, 2020, 9(5): 386-396. [[Crossref](#)]
- Calabrese V, Scapagnini G, Colombrita C, Ravagna A, Penlisi G, Giuffrida Stella AM, et al. Redox regulation of heat shock protein expression in aging and neurodegenerative disorders associated with oxidative stress: a nutritional approach. *Amino Acids*, 2003, 25(3-4): 437-444. [[Crossref](#)]
- Jiménez-Jiménez FJ, Alonso-Navarro H, García-Martín E, & Agúndez JAG. Coenzyme Q10 and dementia: a systematic review. *Antioxidants (Basel)*, 2023, 12(2): 533-543. [[Crossref](#)]
- Chang PS, Chou HH, Lai TJ, Yen CH, Pan JC, & Lin PT. Investigation of coenzyme Q10 status, serum amyloid- β , and tau protein in patients with dementia. *Front Aging Neurosci*, 2022, 14: 910289. [[Crossref](#)]
- Bagheri S, Haddadi R, Saki S, Kourosh-Arabi M, Rashno M, Mojaver A, et al. Neuroprotective effects of coenzyme Q10 on neurological diseases: a review article. *Front Neurosci*, 2023, 17: 1188839. [[Crossref](#)]
- Rauchová H. Coenzyme Q10 effects in neurological diseases. *Physiol Res*, 2021, 70(Suppl4): S683-s714. [[Crossref](#)]
- Abuelezz SA, & Hendawy N. Spotlight on coenzyme Q10 in scopolamine-induced Alzheimer's disease: oxidative stress/PI3K/AKT/GSK 3 β /CREB/BDNF/TrkB. *J Pharm Pharmacol*, 2023, 75(8): 1119-1129. [[Crossref](#)]
- Fišar Z, & Hroudová J. CoQ(10) and mitochondrial dysfunction in Alzheimer's disease. *Antioxidants (Basel)*, 2024, 13(2): 191-203. [[Crossref](#)]
- Vázquez-Jiménez LK, Juárez-Saldivar A, Gómez-Escobedo R, Delgado-Maldonado T, Méndez-Álvarez D, Palos I, et al. Ligand-based virtual screening and molecular docking of benzimidazoles as potential inhibitors of triosephosphate isomerase identified new trypanocidal agents. *Int J Mol Sci*, 2022, 23(17):10047. [[Crossref](#)]
- O'Boyle NM, Morley C, & Hutchison GR. Pybel: a python wrapper for the openbabel cheminformatics toolkit. *Chem Cent J*, 2008, 2: 5-14. [[Crossref](#)]
- Laskowski RA, MacArthur MW, & Thornton JM. Validation of protein models derived from experiment. *Curr Opin Struct Biol*, 1998, 8(5): 631-639. [[Crossref](#)]
- Colovos C, & Yeates TO. Verification of protein structures: patterns of nonbonded atomic interactions. *Protein Sci*, 1993, 2(9): 1511-1519. [[Crossref](#)]
- Wiederstein M, & Sippl MJ. ProSA-web: interactive web service for the recognition of errors in three-dimensional structures of proteins. *Nucleic Acids Res*, 2007, 35(Web Server issue): W407-410. [[Crossref](#)]
- A. Yakubu, Donato MD, & Imumorin IG. Modelling func-

- tional and structural impact of non-synonymous single nucleotide polymorphisms of the DQA1 gene of three Nigerian goat breeds. *S Afr J Anim Sci*, 2017, 47(2): 146–156. [Crossref]
27. Kwofie SK, Dankwa B, Odame EA, Agamah FE, Doe LPA, Teye J, et al. *In silico* screening of isocitrate lyase for novel anti-buruli ulcer natural products originating from Africa. *Molecules*, 2018, 23(7): 1550-1560. [Crossref]
 28. Pettersen EF, Goddard TD, Huang CC, Meng EC, Couch GS, Croll TI, et al. UCSF ChimeraX: structure visualization for researchers, educators, and developers. *Protein Sci*, 2021, 30(1): 70-82. [Crossref]
 29. Goddard TD, Huang CC, Meng EC, Pettersen EF, Couch GS, Morris JH, et al. UCSF ChimeraX: meeting modern challenges in visualization and analysis. *Protein Sci*, 2018, 27(1): 14-25. [Crossref]
 30. Wang S, Jiang JH, Li RY, & Deng P. Docking-based virtual screening of T β R1 inhibitors: evaluation of pose prediction and scoring functions. *BMC Chem*, 2020, 14(1): 52-62. [Crossref]
 31. Alhawarri MB, Dianita R, Rawa MSA, Nogawa T, & Wahab HA. Potential anti-cholinesterase activity of bioactive compounds extracted from *Cassia grandis* L.f. and *Cassia timoriensis* DC. *Plants (Basel)*, 2023, 12(2): 344-354. [Crossref]
 32. McCullagh M, Zeczycki TN, Kariyawasam CS, Durie CL, Halkidis K, Fitzkee NC, et al. What is allosteric regulation? Exploring the exceptions that prove the rule! *J Biol Chem*, 2024, 300(3): 105672. [Crossref]
 33. Pawar RP, & Rohane SH. Role of autodock vina in PyRx molecular docking. *J Res Chem*, 2021, 14(2): 132–134. [Crossref]
 34. E. López-Camacho, M. J. García-Godoy, J. García-Nieto, A. J. Nebro, & J. F. Aldana-Montes. Lecture notes in computer science (including subseries lecture notes in artificial intelligence and lecture notes in bioinformatics). 2016, 97(02): 65-77. [Crossref]
 35. Mir WR, Bhat BA, Rather MA, Muzamil S, Almilaibary A, Alkhanani M, et al. Molecular docking analysis and evaluation of the antimicrobial properties of the constituents of *Geranium wallichianum* D. Don ex Sweet from Kashmir Himalaya. *Sci Rep*, 2022, 12(1): 12547. [Crossref]
 36. Daoud NE, Borah P, Deb PK, Venugopala KN, Hourani W, Alzweiri M, et al. ADMET profiling in drug discovery and development: perspectives of *In silico*, *in vitro* and integrated approaches. *Curr Drug Metab*, 2021, 22(7): 503-522. [Crossref]
 37. V. Ivanović, M. Rančić, B. Arsić, & A. Pavlović. Lipinski's rule of five, famous extensions and famous exceptions. *Chemia Naissensis*, 2020, 3(1): 171–181. [Crossref]
 38. Aarthy M, & Singh SK. Envisaging the conformational space of proteins by coupling machine learning and molecular dynamics. *Adv Protein Mol Struct Biol Metho*, 2022: 467–475. [Crossref]
 39. Priestle JP, & Paris CG. Experimental techniques and data banks. *Guidebook on Molecular Modeling in Drug Design*, 1996: 139–217. [Crossref]
 40. Stielow M, Witczyńska A, Kubryń N, Fijałkowski Ł, Nowaczyk J, & Nowaczyk A. The bioavailability of drugs—the current state of knowledge. *Molecules*, 2023, 28(24): 8038-8048. [Crossref]
 41. Kus M, Ibragimow I, & Piotrowska-Kempisty H. Caco-2 cell line standardization with pharmaceutical requirements and *in vitro* model suitability for permeability assays. *Pharmaceutics*, 2023, 15(11): 2523-2533. [Crossref]
 42. Van Den Abeele J, Rubbens J, Brouwers J, & Augustijns P. The dynamic gastric environment and its impact on drug and formulation behaviour. *Eur J Pharm Sci*, 2017, 96: 207-231. [Crossref]
 43. Neupane R, Boddu SHS, Renukuntla J, Babu RJ, & Tiwari AK. Alternatives to biological skin in permeation studies: current trends and possibilities. *Pharmaceutics*, 2020, 12(2): 152-162. [Crossref]
 44. Nguyen TT, Duong VA, & Maeng HJ. Pharmaceutical formulations with p-glycoprotein inhibitory effect as promising approaches for enhancing oral drug absorption and bioavailability. *Pharmaceutics*, 2021, 13(7): 1103-1113. [Crossref]
 45. Callaghan R, Luk F, & Bebawy M. Inhibition of the multidrug resistance P-glycoprotein: time for a change of strategy? *Drug Metab Dispos*, 2014, 42(4): 623-631. [Crossref]
 46. Holford N, & Yim DS. Volume of distribution. *Transl Clin Pharmacol*, 2023, 24: 74-77. [Crossref]
 47. Dasgupta A. Monitoring free mycophenolic acid concentration: is there any clinical advantage? 2016: 83-107.
 48. Nau R, Sörgel F, & Eiffert H. Penetration of drugs through the blood-cerebrospinal fluid/blood-brain barrier for treatment of central nervous system infections. *Clin Microbiol Rev*, 2010, 23(4): 858-883. [Crossref]
 49. Carpenter TS, Kirshner DA, Lau EY, Wong SE, Nilmeier JP, & Lightstone FC. A method to predict blood-brain barrier permeability of drug-like compounds using molecular dynamics simulations. *Biophys J*, 2014, 107(3): 630-641. [Crossref]
 50. Taylor C, Crosby I, Yip V, Maguire P, Pirmohamed M, & Turner RM. A Review of the important role of CYP2D6 in pharmacogenomics. *Genes (Basel)*, 2020, 11(11): 1295-1302. [Crossref]
 51. Lee J, Beers JL, Geffert RM, & Jackson KD. A review of CYP-mediated drug interactions: mechanisms and *in vitro* drug-drug interaction assessment. *Biomolecules*, 2024, 14(1): 99-101. [Crossref]
 52. Shubbar Q, Alchakee A, Issa KW, Adi AJ, Shorbagi AI, & Saber-Ayad M. From genes to drugs: CYP2C19 and pharmacogenetics in clinical practice. *Front Pharmacol*, 2024, 15: 1326776. [Crossref]
 53. Hacker K, Maas R, Kornhuber J, Fromm MF, & Zolk O. Substrate-dependent inhibition of the human organic cation transporter OCT2: a comparison of metformin with experimental substrates. *PLoS One*, 2015, 10(9): e0136451. [Crossref]
 54. Thomas DN, Wills JW, Tracey H, Baldwin SJ, Burman M, Williams AN, et al. Ames test study designs for nitrosamine mutagenicity testing: qualitative and quantitative analysis of key assay parameters. *Mutagenesis*, 2024, 39(2): 78-95. [Crossref]

55. Gad SC. Maximum tolerated dose. In: *Encyclopedia of Toxicology (Fourth Edition)*. 2024, edn. Academic Press.
56. Stergiopoulos C, Tsopelas F, & Valko K. Prediction of hERG inhibition of drug discovery compounds using biomimetic HPLC measurements. *Admet dmpk*, 2021, 9(3): 191-207. [[Crossref](#)]
57. Niyomchan A, Chatgat W, Chatawatee B, Keereekoch T, Issuriya A, Jaisamut P, *et al.* Safety evaluation of the polyherbal formulation nawatab: acute and subacute oral toxicity studies in rats. *Evid Based Complement Alternat Med*, 2023, 2023: 9413458. [[Crossref](#)]
58. Selvestrel G, Lavado GJ, Toropova AP, Toropov AA, Gadaleta D, Marzo M, *et al.* Monte carlo models for subchronic repeated-dose toxicity: systemic and organ-specific toxicity. *Int J Mol Sci*, 2022, 23(12): 6615-6625. [[Crossref](#)]
59. Björnsson ES. Hepatotoxicity by drugs: the most common implicated agents. *Int J Mol Sci*, 2016, 17(2): 224-234. [[Crossref](#)]
60. Bialas I, Zelent-Kraciuk S, & Jurowski K. The skin sensitisation of cosmetic ingredients: review of actual regulatory status. *Toxics*, 2023, 11(4): 392-404. [[Crossref](#)]
61. Luan F, Wang T, Tang L, Zhang S, & Cordeiro M. Estimation of the toxicity of different substituted aromatic compounds to the aquatic ciliate *Tetrahymena pyriformis* by QSAR approach. *Molecules*, 2018, 23(5): 1002-1013. [[Crossref](#)]
62. Wu X, Zhang Q, & Hu J. QSAR study of the acute toxicity to fathead minnow based on a large dataset. *SAR QSAR Environ Res*, 2016, 27(2): 147-164. [[Crossref](#)]
63. Dumont M, Kipiani K, Yu F, Wille E, Katz M, Calingasan NY, *et al.* Coenzyme Q10 decreases amyloid pathology and improves behavior in a transgenic mouse model of Alzheimer's disease. *J Alzheimers Dis*, 2011, 27(1): 211-223. [[Crossref](#)]
64. Hernández-Camacho JD, Bernier M, López-Lluch G, & Navas P. Coenzyme Q(10) supplementation in aging and disease. *Front Physiol*, 2018, 9: 44-54. [[Crossref](#)]
65. Zhu ZG, Sun MX, Zhang WL, Wang WW, Jin YM, & Xie CL. The efficacy and safety of coenzyme Q10 in Parkinson's disease: a meta-analysis of randomized controlled trials. *Neurol Sci*, 2017, 38(2): 215-224. [[Crossref](#)]
66. Turunen M, Olsson J, & Dallner G. Metabolism and function of coenzyme Q. *Biochim Biophys Acta*, 2004, 1660(1-2): 171-199. [[Crossref](#)]
67. Kitchen DB, Decornez H, Furr JR, & Bajorath J. Docking and scoring in virtual screening for drug discovery: methods and applications. *Nat Rev Drug Discov*, 2004, 3(11): 935-949. [[Crossref](#)]

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